Acute flaccid paralysis associated with enterovirus D68 infection: a case report

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Case report

In July 2017, a 28-month-old boy presented to a private outpatient clinic with a 2-day history of fever and coryzal symptoms (Table). He had enjoyed good past health and his family history was unremarkable. On clinical examination, he was noted to have respiratory distress and tachycardia. Plain radiograph of the chest (CXR) showed perihilar haziness but no consolidation. He was transferred to Queen Mary Hospital, Hong Kong, to exclude myocarditis in view of elevated serum troponin in blood taken in the private clinic. On admission, he was noted to have diffuse crepitations and wheeze suggestive of pneumonitis. Nebulised salbutamol, hypertonic saline, and intravenous cefotaxime were administered. Complete blood count revealed neutrophilia, normal liver and renal function, and normal creatine kinase level. Venous blood gas showed no acidosis. Troponin was high initially but then gradually normalised. Echocardiogram showed no features of myocarditis. Nasopharyngeal aspirate and throat swab test results were positive for enterovirus (EV)/rhinovirus ribonucleic acid (RNA) using reverse transcriptase-polymerase chain reaction (RT-PCR) detection method. EV71 RNA was not evident. Nasopharyngeal aspirate for mycoplasma, throat swab and blood culture were all negative. He then developed progressive respiratory distress with increased cough and high fever up to 40°C on day 7 of illness and CXR showed worsening of bilateral perihilar haziness. In view of the progressive respiratory failure, the child was admitted to the paediatric intensive care unit 2 days later (day 9).

On admission to paediatric intensive care unit, he was noted to have generalised muscle weakness with a weak voice and an inability to sit up. Gag reflex and jerks were preserved. He was then intubated and ventilated under sedation. On reassessment, chest auscultation revealed decreased air entry over the left side, corresponding to the left lower zone collapse evident on CXR. He was prescribed piperacillintazobactam and levofloxacin. On day 10, throat swab culture grew only commensals while endotracheal aspirate revealed scanty growth of alpha-haemolytic streptococci with negative fungal smear and culture, and RT-PCR identified EV/rhinovirus. Bronchoalveolar lavage was also performed but results were unremarkable: *Pneumocystis jiroveci (carinii)*, smear and culture for bacteria, fungus and acid-fast bacilli, and RT-PCR for cytomegalovirus, herpes simplex virus, *Mycoplasma, Legionella*, were all negative. Urine culture was negative for Legionella antigen.

On day 11, the patient exhibited paradoxical breathing on weaning of sedation along with hypotonia and paralysis of four limbs. Urgent sagittal T2-weighted magnetic resonance imaging of spine (day 11) showed mild T2 hyperintensity with mild expansion within the central portion of the cervical cord from C3 to C6 (Fig). No intraspinal mass or collection could be seen. Neurology examination was performed on day 12. Creatine kinase was normal and anti-acetylcholine receptor and antiaquaporin-4 were negative. Eye examination the following day showed no evidence of optic neuritis. Immunoglobulin G and immunoglobulin M of antigangliosides were all negative. Lumbar puncture revealed normal cerebrospinal fluid level of glucose and protein. Total cell count was $100 \times 10^6/L$ with predominantly (80%) neutrophils. Cerebrospinal fluid culture was negative for viruses, including EV, herpes simplex virus or varicella-zoster virus. Cerebrospinal fluid levels of oligoclonal protein and immunoglobulin G were unremarkable. The working diagnosis was transverse myelitis affecting cervical cord C3 to C6. He was prescribed pulse methylprednisolone 30 mg/kg for 5 days (day 12 to 16) followed by a tapering oral dose of prednisolone together with intravenous immunoglobulin 2 g/kg over 2 days (day 16 and 17). He was also treated with therapeutic plasma exchange with 1.5-times plasma volume for five courses over 2 weeks (day 22, 24, 26, 30, 32). His condition gradually improved and he was extubated (day 47) after 38 days of invasive ventilation.

Day from symptom onset	Clinical manifestations	Investigation results	Treatment and management
Day 1	 Fever and coryzal symptoms for 2 days at GOPC in private hospital RD and tachycardia on exam 	CXR: perihilar haziness, no consolidation	
Day 3	 Transferred and admitted to Queen Mary Hospital to rule out myocarditis Haemodynamically stable but noted diffuse crepitations and wheeze suggestive of pneumonitis 	 Initially elevated troponin, subsequently normalised. ECHO: no signs of myocarditis CBC showed neutrophilia LRFT, CK, CK-MB and VBG normal NPA EV/RV RNA RT-PCR positive, EV71 RNA not detected by RT-PCR NPA for mycoplasma, TS and blood culture all negative EV-D68 RNA were detected by gene sequencing subsequently on day 30 in NPA, ETA and stool samples 	 Nebulised salbutamol and nebulised hypertonic saline IV cefotaxime
Day 7	 Progressive RD with increased cough, swinging fever up to 40°C 	• CXR: worsening of bilateral perihilar haziness	
Day 9	 Admitted to PICU in view of progressive respiratory failure 		
Day 10	 Generalised muscle weakness, unable to sit up, weak voice Gag reflex and jerks still present Decreased air entry over left side on chest auscultation revealed, corresponding to CXR finding of left lower zone collapse 	 TS culture: commensals only ETA: EV/RV PCR positive, scanty growth of alpha-haemolytic streptococci, negative fungal smear and culture BAL: PCP, bacterial, AFB and fungal smear and culture all negative. <i>Mycoplasma</i>, <i>Legionella</i>, CMV, HSV PCR all negative Urine for Legionella Ag negative 	 Sedated, intubated and ventilated IV piperacillin-tazobactam and levofloxacin
Day 11	 Increased paradoxical breathing movement on weaning down sedation, also developed AFP with, hypotonia and paralysis of four limbs. Jerks still present with downgoing plantars. Sensation could not be tested as child was too weak to communicate 	 Normal CK Urgent sagittal T2-weighted MRI spine showed mild T2 hyperintensity with mild expansion within central portion of cervical cord (C3-C6). No intraspinal mass or collection could be seen. Overall features were suggestive of ATM 	
Day 12	 Working diagnosis was ATM affecting cervical cord C3-C6 levels. LP was performed with no evidence of NMO, MS or GBS Subsequently established definitive diagnosis of EV-D68-associated AFP on day 30 	 AChR and anti-aquaporin-4[†] negative Eye exam: no evidence of optic neuritis IgG and IgM of anti-gangliosides[‡]: negative LP showed normal CSF glucose and protein. TCC 100 × 10⁶/L with neutrophil predominance (80%). CSF culture negative. EV, HSV, VZV all not detected CSF oligoclonal protein[§] was negative and CSF IgG^{II} level was unremarkable EV-D68 was not demonstrated in CSF sample and plasma/serum sample was not tested for EV-D68 PCR 	 Pulse MP 30 mg/kg for 5 days (day 12-16) followed by tapering dose of oral prednisolone IVIG 2 gm/kg over 2 days (day 16 and 17) TPE (1.5-times plasma volume) for 5 courses over 2 weeks (day 22, 24, 26, 30, 32)
Day 47	 Successfully extubated after total 38 days of invasive ventilation 	 Repeated NPA, ETA and stool on day 30 remained positive for EV-D68 which then turned negative on day 55 	
Day 64	 Successfully discharged from PICU after 2 months' stay Able to achieve full neurological recovery with intensive training by physiotherapists 		

Abbreviations: AChR = anti-acetylcholine receptor; AFB = acid-fast bacilli; AFP = acute flaccid paralysis; Ag = antigen; ATM = acute transverse myelitis; BAL = broncho-alveolar lavage; CBC = complete blood count; CK = creatine kinase; CK-MB = creatine kinase in muscle/brain; CMV = cytomegalovirus; CSF = cerebrospinal fluid; CXR = chest X-ray; ECHO = echocardiogram; ETA = endotracheal aspirate; EV = enterovirus; GBS = Guillain–Barré syndrome; GOPC = general out-patient clinic; HSV = herpes simplex virus; IgG = immunoglobulin G; IgM = immunoglobulin M; IV = intravenous; IVIG = intravenous immunoglobulin; LP = lumbar puncture; LRFT = liver and renal function tests; MFS = Miller–Fisher syndrome; MND = motor neuron disease; MP = methylprednisolone; MRI = magnetic resonance imaging; MS = multiple sclerosis; NMO = neuromyelitis optica; NPA = nasopharyngeal aspirate; PCP = pneumocystis pneumonia (jiroveci); PCR = polymerase chain reaction; PICU = paediatric intensive care unit; RD = respiratory distress; RNA = ribonucleic acid; RT-PCR = reverse transcriptase-polymerase chain reaction; RV = rhinovirus; TCC = total cell count; TPE = therapeutic plasma exchange; TS = throat swab; VBG = venous blood gas; VZV= varicella-zoster virus

[†] Anti-aquaporin-4 is a sensitive and highly specific serum marker of NMO

[±] Anti-ganglioside is panel used for diagnosis of motor neuropathies (GBS, MND, MFS, and multifocal motor neuropathies)

[§] CSF oligoclonal protein is positive in 80% of patients with MS

CSF IgG index is elevated in 80% of patients with MS

The child was successfully discharged from the paediatric intensive care unit after 2 months (day 64) and achieved full neurological recovery with intensive training by physiotherapists. Subsequent review by a microbiologist using gene sequencing of initial specimens obtained on day 3 of admission to Queen Mary Hospital revealed EV-D68 RNA in nasopharyngeal aspirate, endotracheal aspirate and stool samples. Samples remained positive for 4 weeks (day 30, during acute deterioration warranting intensive care unit admission) and were negative after 6 weeks (day 55). The definitive diagnosis was EV-D68-associated acute flaccid paralysis (AFP) although EV-D68 was not present in the cerebrospinal fluid and plasma/serum samples were not tested for EV-D68 by RT-PCR.

Discussion

Acute flaccid paralysis is defined by the World Health Organization as a clinical syndrome of diverse aetiology characterised by acute-onset limb weakness or paralysis with varying degrees of autonomic and somatic nervous system dysfunction that reaches maximum severity over a period of days or weeks in a child younger than 15 years of age.¹ It is a diagnosis of exclusion. In 1962, a new strain of EV, EV-D68, was identified in Berkeley, California. In 2014, EV-D68 outbreaks were reported in 20 countries including the United States, Canada, Europe, and Asia with a total of over 2000 cases. This corresponded to an increased global incidence of AFP.² A casual association between EV-D68 and AFP is supported by Bradford Hill criteria.³ Despite public health attempts in 1988 to eliminate AFP through the Global Polio Eradication Initiative⁴ and roll-out of the oral polio vaccine⁵ to prevent vaccine-associated poliomyelitis, the emergence of EV-D68-associated AFP has become a significant cause of neurological deficits in children since 2014. Owing to its impact on the healthcare system, a comprehensive literature review and further detailed studies are warranted. This is the first case encountered in our department. It is important for clinicians in Hong Kong to be alert for the disease.

As a newly emerging disease manifestation, a high index of suspicion and clinical awareness is advocated to facilitate earlier recognition and diagnosis through appropriate investigations, and presumably, improved clinical outcomes. The optimum treatment strategy has yet to be defined and preventive strategies are still being developed. Local and international notification systems as well as comprehensive surveillance are suggested since disease outbreaks may occur at any time and may have a serious impact on affected children.

Author contributions

Concept or design: WYK Chan, PL Ho.

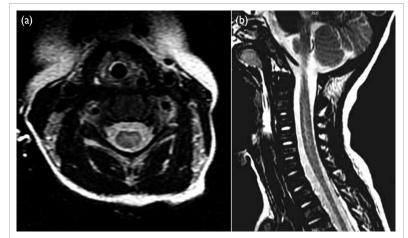


FIG. Axial (a) and sagittal (b) images on T2-weighted magnetic resonance imaging of spine showing mild T2 hyperintensity with mild expansion within central portion of cervical cord from C3 to C6

Acquisition of data: WYK Chan, SHY Chim, DML Tse. Analysis or interpretation of data: WYK Chan, DML Tse, PL Ho.

Drafting of the manuscript: WYK Chan.

Critical revision of the manuscript for important intellectual content: All authors.

All authors had full access to the data, contributed to the study, approved the final version for publication, and take responsibility for its accuracy and integrity.

Conflicts of interest

All authors have disclosed no conflicts of interest.

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Ethics approval

The parents of the patient gave consent for publication.

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