ABSTRACT

Introduction: There is a global trend of increasing macrolide and fluoroquinolone resistance in *Mycoplasma genitalium* (MG), such that international guidelines recommend molecular detection of resistance if a patient has MG-positive test results. Tests for MG are not routinely performed in Hong Kong. This study examined the detection of MG in endocervical swabs and the associated macrolide and fluoroquinolone resistance rates.

Methods: Endocervical swabs received from two sexual health clinics in Hong Kong for routine assessments of *Chlamydia trachomatis* and *Neisseria gonorrhoeae* were also subjected to detection of MG. All MG-positive samples were tested for resistance-mediating mutations in 23S rRNA, *parC*, and *gyrA* genes. Laboratory records and past results for each patient were analysed.

Results: In total, endocervical swabs from 285 patients were included in this study. *Mycoplasma genitalium* was detected in swabs from 21 patients (7.4%) by real-time polymerase chain reaction with a commercial kit. Among MG-positive samples which were successfully analysed further, macrolide resistance-mediating mutations in 23S rRNA were found in 42.1% (8/19); fluoroquinolone resistance–related mutations in *parC* and *gyrA* were found in 65% (13/20) and 0% (0/20), respectively. All macrolide-resistant MG strains were also fluoroquinolone-resistant (42.1%, 8/19). No assessed factors were associated with the detection of MG or resistance-related mutations.

Conclusion: In Hong Kong, MG was detected in endocervical swabs from 7.4% of patients in sexual health clinics, with high rates of macrolide and fluoroquinolone resistance. These findings warrant careful review of testing, clinical correlation, and treatment strategies for MG in the context of increasing antibiotic resistance.

New knowledge added by this study

- *Mycoplasma genitalium* (MG) was detected in endocervical swabs from 7.4% of patients in two sexual health clinics in Hong Kong.
- High rates of macrolide and fluoroquinolone resistance–associated mutations (42.1% and 65%, respectively) were detected in MG-positive specimens.
- All macrolide-resistant MG strains were also fluoroquinolone-resistant.
- No clinical or demographic factors were significantly associated with the detection of MG or resistance-related mutations.

Implications for clinical practice or policy

- Our findings support the existing recommendation that testing should be reserved only for patients with increased risks or for whom treatment has failed, as well as their contacts.
- The empirical uses of macrolide and fluoroquinolone regimens utilised in Hong Kong might explain the high rates of resistance found in this study.
- Careful review is required with respect to testing, clinical correlation, and treatment strategies for MG in the context of increasing antibiotic resistance.

Introduction

*Mycoplasma genitalium* (MG) is capable of causing urogenital infection, especially urethritis, in men. There is increasing evidence to support its ability to cause cervicitis and pelvic inflammatory disease in women.\(^1\) Data from a recent meta-analysis showed a prevalence of 1.3% in the general populations of developed countries, with similar rates among
Methods

Specimen and data collection

From March to May 2019, endocervical swabs of female patients sent from two STI clinics in Hong Kong for routine molecular detection of *Chlamydia trachomatis* (CT) and *Neisseria gonorrhoeae* (NG) were also subjected to detection of MG. Information regarding the presence or absence of genitourinary symptoms was provided by attending physicians. All available laboratory information for each tested patient was reviewed to determine demographic data, history of STI, human immunodeficiency virus status, and test results from the current visit.

Detection of *Mycoplasma genitalium*

The cobas TV/MG assay (Roche Diagnostics, Rotkreuz, Switzerland) performed on the cobas 6800 System (Roche Diagnostics) was used for detection of MG. The cobas TV/MG assay is a CE-marked, Food and Drug Administration–cleared, commercial qualitative nucleic acid test that utilises the real-time polymerase chain reaction for dual detection of *Trichomonas vaginalis* (TV) and MG. Its reported overall sensitivity and specificity rates were 83.1% and 98.4%, respectively, for MG detection in endocervical swabs from female patients. Tests were performed in accordance with the manufacturer’s instructions.

Detection of other sexually transmitted pathogens

In brief, CT and NG were detected by the cobas CT/NG assay (Roche Diagnostics), while herpes simplex virus (HSV) 1 and HSV 2 were detected by the cobas HSV 1 and 2 assay (Roche Diagnostics), both performed on the cobas 4800 System (Roche Diagnostics). In addition, NG was detected by the culture method on modified Thayer-Martin agars. Either molecular– or culture-based test indicating the presence of NG was considered a positive result. *Trichomonas vaginalis* was detected by the cobas TV/MG assay (Roche Diagnostics) and wet mount microscopy after enrichment in Feinberg medium. Similar to NG, either test indicating the presence of TV was considered a positive result. Syphilis was detected by serological tests of serum specimens, including enzyme immunoassay (DiaSorin, Saluggia, Italy), Venereal Disease Research Laboratory test (Becton, Dickinson and Company, Franklin Lakes [NJ], US), fluorescent treponemal antibody absorption test (ImmunoDiagnostics Limited, Hong Kong, China), or *Treponema pallidum* passive particle agglutination test (Fujirebio Diagnostics AB, Göteborg, Sweden).
Detection of macrolide and fluoroquinolone resistance

Mutations at nucleotide positions 2071 and 2072 (2058 and 2059, respectively, by Escherichia coli numbering) of the 23S rRNA gene have been associated with macrolide resistance in MG and subsequent treatment failure; mutations in the quinolone resistance-determining region of the parC gene, and possibly the gyrA gene, have been associated with FQ resistance. Polymerase chain reaction analysis of these genes was performed as described previously. Sequencing was performed using a 3730xl DNA Analyzer (Applied Biosystems, Foster City [CA], US), in accordance with the manufacturer’s instructions. Primers used for sequencing were the same as those used for the polymerase chain reaction. Resulting sequences were compared to the sequence of wild-type strain MG G37 by using BLAST (https://blast.ncbi.nlm.nih.gov/Blast.cgi).

Statistical analysis

Calculation of odds ratios, and the Chi squared test or Fisher’s exact test, were performed as univariate analysis to identify associations between assessed factors (ie, age, previous STI clinic visit, history of STI, symptoms, and sexually transmitted co-infections) and outcomes (ie, detection of MG and detection of resistance mutations), as well as between STI and symptoms. Because of the low outcome frequency, Firth logistic regression was employed to analyse associations between factors with P<0.25 in univariate analysis and the detection of resistance mutations. IBM SPSS Statistics Subscription (Windows version, IBM Corp, Armonk [NY], US) was used for data analysis.

The STROBE statement reporting guidelines were followed in this study.

Results

In total, 285 non-duplicated specimens from 285 patients were included in this study. The mean patient age was 35.5 years (range, 16-76 years); 23.9% of the patients (n=68) were in the younger age-group (≤25 years). Of the 285 patients, 59.6% (n=170) were new patients without a previous relevant testing record. At the low outcome frequency, Firth logistic regression was employed to analyse associations between factors with P<0.25 in univariate analysis and the detection of resistance mutations. IBM SPSS Statistics Subscription (Windows version, IBM Corp, Armonk [NY], US) was used for data analysis.

The STROBE statement reporting guidelines were followed in this study.

Younger age, previous STI clinic visit, history of STI, symptoms, and sexually transmitted co-infections were not associated with the detection of MG (Table 2). Symptoms were only significantly associated with the detection of CT (Table 3).

Among the 21 MG-positive endocervical swabs, sequencing results for 23S rRNA were available for 19 specimens. In total, 42.1% of the specimens (n=8) harboured the macrolide resistance–mediating mutations A2071G or A2072G. Regarding parC and gyrA, sequencing results were available for 20 specimens. Overall, 65% of the specimens (n=13) harboured the FQ resistance–related mutations G248T (Ser83Ile), G259T (Asp87Tyr), or G259A (Asp87Asn) within parC; no mutations were detected in gyrA. Among the 19 specimens with sequencing results available regarding both macrolide and FQ resistance, dual resistance was detected in 42.1% of the specimens (n=8); thus, all macrolide-resistant strains were also resistant to FQ. Furthermore, C184T (Pro62Ser) in parC, for which clinical significance is unknown, was detected in one specimen without any other mutation and in one specimen with dual resistance.

In subgroup analysis of patients with MG-positive specimens, younger age, previous STI clinic visit, symptoms, and sexually transmitted co-infections were not associated with the detection of resistance mutations, similar to the findings among all specimens; however, a history of STI was

<table>
<thead>
<tr>
<th>Sexually transmitted infection</th>
<th>No. of patients with positive results</th>
<th>No. of symptomatic patients with positive results</th>
</tr>
</thead>
<tbody>
<tr>
<td>MG</td>
<td>14</td>
<td>10</td>
</tr>
<tr>
<td>MG, CT</td>
<td>5</td>
<td>4</td>
</tr>
<tr>
<td>MG, TV</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>CT</td>
<td>27</td>
<td>22</td>
</tr>
<tr>
<td>TV</td>
<td>15</td>
<td>8</td>
</tr>
<tr>
<td>SYP</td>
<td>11</td>
<td>3</td>
</tr>
<tr>
<td>NG</td>
<td>4</td>
<td>3</td>
</tr>
<tr>
<td>HSV 1 or 2</td>
<td>4</td>
<td>3</td>
</tr>
<tr>
<td>CT, TV</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>NG, SYP</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>CT, NG</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>NG, TV</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>CT, NG, TV</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>Negative</td>
<td>193</td>
<td>108</td>
</tr>
<tr>
<td>Total</td>
<td>285</td>
<td>173</td>
</tr>
</tbody>
</table>

Abbreviations: CT = Chlamydia trachomatis; HSV = herpes simplex virus; MG = Mycoplasma genitalium; NG = Neisseria gonorrhoeae; SYP = syphilis; TV = Trichomonas vaginalis

TABLE 1. Sexually transmitted infections detected among 285 patients in Hong Kong

In subgroup analysis of patients with MG-positive specimens, younger age, previous STI clinic visit, symptoms, and sexually transmitted co-infections were not associated with the detection of resistance mutations, similar to the findings among all specimens; however, a history of STI was
negatively associated with the detection of mutations (Table 2). This association did not remain after multivariable logistic regression (odds ratio=0.151, 95% confidence interval=0.004-2.983, P=0.221).

Discussion

In this study, 7.4% of endocervical swabs from women attending STI clinics exhibited MG-positive results, although no assessed factors were obviously associated with the detection of MG. *Mycoplasma genitalium*–positive rates did not significantly differ (P=0.131) among patients who were symptomatic (9.2%) and those who were asymptomatic (4.5%). These rates were comparable with the findings of the aforementioned 2008 cross-sectional study in Hong Kong involving male patients with STI,9 as well as with the findings of a multicentre clinical study in the US.15 Reported rates of MG detection have exhibited considerable variability. A 2018 systematic review2 revealed that higher rates were prevalent among at-risk groups (eg, commercial sex workers and men who have sex with men), in clinic-based settings, and in countries with lower economic development. In Australia, the prevalence of MG was found to range from 2.1% to 13%, depending on the population tested; while a higher prevalence of approximately 15% has been reported in Japan.17 Nevertheless, the prevalence in the general population and asymptomatic patients remained low (1.3%),2 which did not support universal screening. In the present study, symptoms were not associated with the detection of MG. Considering the organism’s uncertain clinical significance and natural history, it is necessary to balance the need to test and the risks of unnecessary treatment, including potential aggravation of antibiotic resistance. We agree with the existing recommendation that testing should be carefully selected, reserved only for patients with increased risks or for whom treatment has failed, as well as their contacts.18

Importantly, the choice of specimen might affect the detection of MG. A recent prospective, multicentre study showed that self or clinician-collected vaginal swabs exhibited the best sensitivities (92%-98.9%), while urinary and endocervical swabs were less sensitive (81.5% and 77.8%, respectively).15 These findings were consistent with the results of prior studies19-21; notably, some studies found that endocervical swabs were more sensitive than urinary specimens,20,21 presumably because of the lower bacterial load in urine.22 Endocervical swabs are the routine specimens sent to our laboratory from STI clinics for molecular detection of CT and NG; we perform assays for detection of MG and CT, as recommended by European guidelines.6 Of note, the relatively low sensitivity of the test with respect to endocervical swabs might also have underestimated the prevalence of MG in our study.

Resistance-related mutations in 23S rRNA and *parC* genes were detected in 42.1% and 65% of MG-positive samples, respectively, among which none harboured mutations in *gyrA*. All macrolide-resistant strains were also FQ-resistant (42.1%). Although the populations have differed among studies, similarly high rates of macrolide resistance.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Detection of MG</th>
<th>Detection of resistance mutations in MG–positive specimens</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Odds ratio (95% CI)</td>
<td>P value</td>
</tr>
<tr>
<td>Age ≤25 years</td>
<td>0.997 (0.351-2.83)</td>
<td>0.996</td>
</tr>
<tr>
<td>Previous STI clinic visit</td>
<td>0.903 (0.362-2.253)</td>
<td>0.827</td>
</tr>
<tr>
<td>Previous STI</td>
<td>1.371 (0.479-3.922)</td>
<td>0.555</td>
</tr>
<tr>
<td>Symptomatic</td>
<td>2.181 (0.776-6.132)</td>
<td>0.131</td>
</tr>
<tr>
<td>Co-infection</td>
<td>1.359 (0.527-3.505)</td>
<td>0.524</td>
</tr>
</tbody>
</table>

Abbreviations: 95% CI = 95% confidence interval; MG = *Mycoplasma genitalium*; STI = sexually transmitted infection

## TABLE 2. Univariate analysis of risk factors for detection of *Mycoplasma genitalium* and detection of resistance mutation in *Mycoplasma genitalium*–positive specimens

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Presence of symptoms</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Odds ratio (95% CI)</td>
</tr>
<tr>
<td>Age ≤25 years</td>
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Abbreviations: 95% CI = 95% confidence interval; CT = *Chlamydia trachomatis*; HSV = herpes simplex virus; MG = *Mycoplasma genitalium*; NG = *Neisseria gonorrhoeae*; SYP = syphilis; TV = *Trichomonas vaginalis*

## TABLE 3. Association between sexually transmitted infections and symptoms

<table>
<thead>
<tr>
<th>Sexually transmitted infection</th>
<th>Presence of symptoms</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Odds ratio (95% CI)</td>
</tr>
<tr>
<td>MG</td>
<td>2.181 (0.776-6.132)</td>
</tr>
<tr>
<td>CT</td>
<td>4.099 (1.618-9.937)</td>
</tr>
<tr>
<td>TV</td>
<td>1.424 (0.526-3.611)</td>
</tr>
<tr>
<td>SYP</td>
<td>0.467 (0.158-1.384)</td>
</tr>
<tr>
<td>NG</td>
<td>6.81 (0.86-54)</td>
</tr>
<tr>
<td>HSV 1 or 2</td>
<td>1.959 (0.201-19.07)</td>
</tr>
</tbody>
</table>

Abbreviations: 95% CI = 95% confidence interval; CT = *Chlamydia trachomatis*; HSV = herpes simplex virus; MG = *Mycoplasma genitalium*; NG = *Neisseria gonorrhoeae*; SYP = syphilis; TV = *Trichomonas vaginalis*
have been reported, while rates of FQ and dual resistance have varied among regions (Table 4).3,23-28 Several studies also demonstrated consistent increases in resistance rates over time.3,28,29 All mutations detected in this study have been described previously; while C184T (Pro62Ser) in \textit{parC} is of unknown significance, others are known to confer antibiotic resistance leading to higher minimal inhibitory concentrations and treatment failure.5,11,12,23 In particular, the extent of FQ resistance is reportedly related to the presence of concurrent \textit{parC} and \textit{gyrA} mutations. Although MG strains with lone \textit{parC} mutations had reduced susceptibilities to FQ, they were able to be eradicated by sitafloxacin and (possibly) moxifloxacin. However, concurrent \textit{gyrA} mutations have been shown to further increase the FQ minimal inhibitory concentrations, leading to treatment failure.23,30

No assessed factors were significantly associated with the detection of MG in this study, possibly due to the limited number of positive samples. The authors of other studies have suggested that a syndromic approach (ie, management of a patient whereby a syndrome is used as a basis for the treatment of the causative organisms) and the use of a single dose of azithromycin for treatment of NG (as part of dual therapy), non-gonococcal urethritis/non-specific genital tract infection, or known MG infection contribute to the emergence of macrolide resistance in MG, because this regimen is suboptimal and might exert selective pressure on resistant strains.28,29,31 A similar phenomenon has been observed with respect to FQ, especially in Japan, where frequent use of the second-line antibiotic sitafloxacin caused selection of resistant strains, leading to high rates of FQ resistance in MG.3 In public clinics in Hong Kong, a single dose of azithromycin or a 1-week course of doxycycline is used as empirical treatment for non-gonococcal urethritis or non-specific genital tract infection. If no culprit pathogen is identified and the patient complains of persistent symptoms during follow-up, a 1-week course of moxifloxacin for possible MG is considered, following exclusion of other causes (eg, non-compliance). These empirical uses of macrolide and FQ regimens might explain the high rates of resistance found in this study. For other regions with lower rates of FQ resistance, the relationship between FQ use and emergence of its resistance in MG requires further investigation.

In the context of increasing drug resistance, international guidelines have suggested follow-up molecular testing for resistance determinants in MG-positive specimens.6-8 In particular, the most recent British and Australian guidelines include revised treatment regimens, which suggest 1 week of doxycycline followed by 3 days of azithromycin as treatment for macrolide-sensitive (or susceptibility unknown) MG, or followed by 7 to 10 days of moxifloxacin as treatment for macrolide-resistant MG.7,8 Alternative antibiotics that might be effective (eg, pristinamycin) require further evaluation.32,33 When the detection of MG and its drug resistance profile is considered after patient selection and careful review of clinical indications, testing at a private laboratory may be sought, because this service is not readily available in the public sector in Hong Kong. Our findings of high macrolide and FQ resistance rates in MG implied that the use of azithromycin and moxifloxacin as empirical first- and second-line therapies, respectively, might be ineffective; furthermore, this approach could induce greater drug resistance. These findings should be taken into consideration in future assessments of treatment guidelines for Hong Kong. The acquisition of updated treatment strategies from international guidelines may also be useful.

Because our laboratory is a reference laboratory
that serves all public STI clinics in Hong Kong, our database is comprehensive in terms of laboratory testing records. Specimens in this study were unique and not duplicated for any patient. In addition, we have considerable capacity to perform arrays of confirmatory tests for various sexually transmitted pathogens. However, this study was limited by the absence of other clinical information such as sexual practices, antimicrobials prescribed, and treatment outcomes, because these data were not available to the authors. The small number of MG-positive samples also limited our ability to assess correlations with factors considered in this study.

**Conclusion**

*Mycoplasma genitalium* was detected in 7.4% of 285 endocervical swabs collected from both symptomatic and asymptomatic women attending STI clinics in Hong Kong. Among the MG-positive samples, macrolide resistance-mediating mutations and fluoroquinolone resistance–related mutations were detected in 42.1% and 65%, respectively. Dual resistance was also detected in all macrolide-resistant strains (42.1%). These findings suggest that both testing and treatment strategies require careful review to avoid further enhancing the prevalence of antibiotic resistance.

**Author contributions**

Concept or design: KKM Ng, PKL Leung.
Analysis of data: All authors.
Drafting of the manuscript: KKM Ng.
Critical revision of the manuscript for important intellectual content: All authors.

All authors had full access to the data, contributed to the study, approved the final version for publication, and take responsibility for its accuracy and integrity.

**Conflicts of interest**

All authors have disclosed no conflicts of interest.

**Acknowledgement**

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**Funding/support**

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**Ethics approval**

This study obtained ethics approval (Ref LM 424/2019) from the Ethics Committee of the Department of Health, Hong Kong SAR Government. Patients consented to testing for sexually transmitted pathogens.

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