The C282Y mutation of the *HFE* gene is not found in Chinese haemochromatotic patients: multicentre retrospective study

WMS Tsui, PWY Lam, KC Lee, KF Ma, YK Chan, MWY Wong, SP Yip, CSC Wong, ASF Chow, STH Lo

Objective. To detect two novel mutations (C282Y and H63D) of the *HFE* gene in Chinese patients with hepatic iron overload.

Design. Multicentre retrospective study.

Setting. Four public hospitals, Hong Kong.

Participants. Fifty Chinese patients who presented from January 1987 through December 1999 with hepatic iron overload from various causes.

Main outcome measures. The DNA from liver biopsy samples was tested for *HFE* mutations by restriction fragment length polymorphism analysis.

Results. The sample DNA quality was unsatisfactory for analysis of the C282Y mutation in one case and the H63D mutation in nine cases. The C282Y mutation was not detected in any of the 49 satisfactory samples. Three of the 41 samples were heterozygous for the H63D mutation and only one was homozygous, giving an allele frequency of 6.1%. Of the three H63D-heterozygotes, one had β -thalassaemia major, one had β -thalassaemia major, one had hereditary spherocytosis. None of the 12 patients who were presumed to have primary haemochromatosis were positive for either mutation.

Conclusions. The classical form of human leukocyte antigen–linked hereditary haemochromatosis appears to be absent from this locality. The H63D mutation is found in a minority (9.8%) of the patients, in whom it may act synergistically with an erythropoietic factor.

HKMJ 2000;6:153-8

Key words: Genes, MHC class I; Hemochromatosis/genetics; Hong Kong; Iron overload/genetics; Mutation, missense

Department of Pathology, Caritas Medical Centre, Kowloon, Hong
Kong
WMS Tsui, FRCPath, FHKAM (Pathology)
ASF Chow, BSc
STH Lo, MSc
Department of Pathology, Queen Elizabeth Hospital, Kowloon,
Hong Kong
PWY Lam, FRCPA, FHKAM (Pathology)
CSC Wong, MAppSc
Department of Pathology, Princess Margaret Hospital, Kowloon,
Hong Kong
KC Lee, FRCPath, FHKAM (Pathology)
Department of Pathology, Alice Ho Nethersole Hospital, Tai Po,
Hong Kong
KF Ma, FRCPath, FHKAM (Pathology)
Department of Medicine, Caritas Medical Centre, Kowloon,
Hong Kong
YK Chan, MMed, FHKAM (Medicine)
Department of Nursing and Health Sciences, Hong Kong Polytechnic
University, Kowloon, Hong Kong
MWY Wong, MPhil, BSc
SP Yip, PhD, MPhil
-
Correspondence to: Dr WMS Tsui

Introduction

In western countries, iron overload is regarded as a problem that predominantly affects the white population in the form of autosomal recessive human leukocyte antigen (HLA)-linked hereditary haemochromatosis (HHC), which has a prevalence of nearly 0.45%.¹ A major breakthrough in the understanding of HHC was the cloning of the causative gene (originally designated HLA-H and now renamed HFE) found on chromosome 6p.² The gene product is an HLA class I-like protein, which is cell membrane-bound and has been reported to interact with the transferrin receptor to regulate cellular iron uptake.^{3,4} A homozygous missense mutation that leads to an amino acid substitution of cysteine by tyrosine at position 282 (C282Y) has been detected in the gene in 83% of 178 American haemochromatosis patients.² Although the frequency

of this mutation has been shown to vary in three continents (North America, northern Europe, and Australia), there is now general agreement that 80% to 100% of HHC patients are homozygous for the C282Y mutation.⁵⁻⁸ A second missense mutation that results in a substitution of histidine by aspartic acid at codon 63 (H63D) has also been identified. This mutation acts synergistically with C282Y in 3% to 5% of the patients, and iron overload can occur in H63D homozygotes.⁹⁻¹¹

In the Chinese population, severe iron overload is mainly associated with Cooley's anaemia, and primary haemochromatosis appears to be a rare disease. The C282Y mutation has been found to be absent in the general population of China.^{12,13} The occurrence of the C282Y mutation among Chinese haemochromatotic patients, however, has not been examined. This study aimed to find the frequency of the C282Y and H63D mutations of *HFE* gene in a group of Chinese patients with iron overload.

Materials and methods

Patients whose liver biopsy results showed increased iron deposition were identified from the computer records from January 1987 through December 1999 of four participating hospitals: the Alice Ho Nethersole Hospital, Caritas Medical Centre, Princess Margaret Hospital, and Queen Elizabeth Hospital. Histological sections of the liver were retrieved and reviewed independently by at least two pathologists. The criterion for inclusion of patients in this study was the presence of a minimum of grade 2 hepatocellular iron, which was assessed in sections that had been stained with Perls' reagent, according to a grading system (on a scale of 0 to 4) modified from Scheuer's method (Table 1).^{14,15} Grade 2 iron corresponds to a moderate degree of iron deposition, whereas grades 3 and 4 indicate severe iron overload equivalent to hepatic iron concentrations of 130 to 850 µmol/g dry weight. According to such criteria, patients with the classical form of HHC should have been included. Formalin-fixed, paraffin-embedded liver biopsy samples were analysed for the C282Y and H63D mutations using polymerase chain reaction (PCR) amplification followed by restriction fragment length polymorphism (RFLP) analysis.

DNA extraction and analysis

Tissue sections from each sample were deparaffinised by washing with three changes of xylene, followed by two changes of absolute ethanol, and one change of 95% alcohol. The tissue fragments were incubated with

 Table 1. Histological grading of hepatocellular iron accumulation^{14,15}

Grade	Haemosiderin deposition in hepatocytes
0	Absent
1	<25% of hepatocytes: granules barely discernible
2	25%-75% of hepatocytes: discrete granules in zone 1 and 2
3	75%-100% of hepatocytes: coarse granular deposit with acinar gradient
4	100% of hepatocytes: massive deposit with obliteration of acinar gradient

proteinase K at 60°C overnight. An equal volume of a solution of phenol, chloroform, and isoamyl alcohol (in a volume ratio of 25:24:1) was added, and the mixture was vortexed thoroughly and incubated on ice for 15 minutes. The mixture was then centrifuged at 13000 rpm for 10 minutes, and the upper aqueous layer containing the DNA was transferred to a new tube. An equal volume of a solution of phenol, chloroform, and isoamyl alcohol was added and the DNA purification procedure was repeated. Two microlitres of SeeDNA (Pharmacia Amersham Biotech, Uppsala, Sweden), and 40 µL of 3 M sodium acetate, and 1 mL of absolute alcohol were added to the DNA solution. The mixture was vortexed briefly and centrifuged at 13000 rpm for 5 minutes. The DNA pellet was washed with 1 mL of 70% alcohol and then 1 mL of absolute alcohol. The DNA was vacuum-dried for 15 minutes and resuspended in 50 µL distilled water.

To detect C282Y and H63D mutations, PCR analysis was performed according to published methods.^{16,17} The primers for the detection of the C282Y mutation were: (forward) 5'-GGA AGA GCA GAG ATA TAC GT-3' and (reverse) 5'-CTC AGG CAC TCC TCT CAA CC-3'. The primers for the detection of the H63D mutation were: (forward) 5'-ACA TGG TTA AGG CCT GTT GC-3' and (reverse) 5'-GCC ACA TCT GGC TTG AAA TT-3'. The cycling conditions for both reactions were as follows: heat activation, 10 minutes; 40 cycles of incubation of 94°C for 1 minute (denaturation), 54°C for 1 minute (reannealing), and 72°C for 1 minute (extension); and an extra 7 minutes' extension for the last cycle. The sizes of the PCR products bearing the C282Y and H63D mutations were 392 bp and 208 bp, respectively (Fig). Subsequent digestion of the C282Y-mutated PCR product with the restriction enzyme SnaB1 yielded 111-bp and 281-bp fragments of the mutant allele; the wild-type allele was not digested. In contrast, the PCR product containing the H63D mutation yielded 70-bp and 138-bp fragments for the normal allele after digestion with NdeII; the mutant allele was not digested.

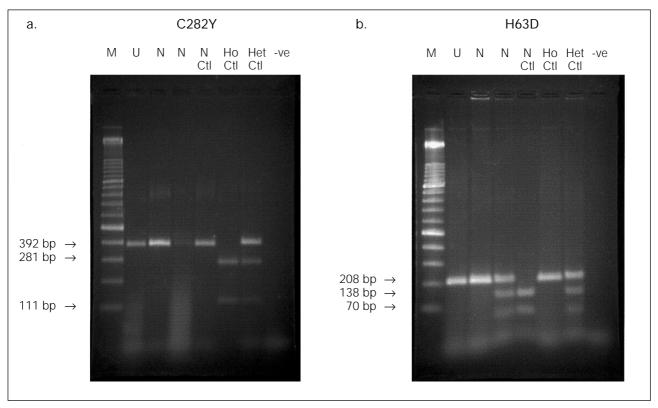


Fig. Example of agarose gel of electrophoresed amplification products to detect C282Y and H63D mutations Samples were incubated with *Sna*BI or *Nde*II to detect (a) C282Y and (b) H63D mutation, respectively; M = 100-bp molecular weight markers, U = undigested, N = homozygous wild-type, Ho = homozygous mutant, Het = heterozygous, Ctl = control, -ve = negative control

Results

A total of 50 patients (27 males and 23 females) were studied. All patients were Chinese; their mean age was 46.2 years (range, 9-88 years). Their medical records (available in 39 cases) and laboratory data were reviewed, and pertinent clinical data were analysed. According to histological assessment and clinical correlation, the majority of the cases of iron overload were cases of secondary haemochromatosis, which could be divided into the following four disease groups: (1) Cooley's anaemia requiring regular blood transfusion (n=7); (2) haemoglobinopathies or corpuscular abnormalities not requiring transfusion (n=9); (3) alcoholic liver disease and cirrhosis of various causes (n=9); and (4) miscellaneous diseases (eg leukaemia, malignant tumours) requiring occasional transfusion (n=13). A fifth group of patients had idiopathic iron overload and had been regarded from clinical findings as having primary haemochromatosis (n=12). The diagnosis in this group was made because of the combination of an elevated serum ferritin level and iron saturation, compatible histological findings, at least grade 2 siderosis, and an absence of an obvious secondary cause. The family histories of iron overload in this group of patients were not available; their characteristics are summarised in Table 2.

Table 2. Characteristics of patients categorised as
having idiopathic haemochromatosis

Characteristic	Value
Sex ratio (M:F)	9:3
Age (years)	
Mean (range)	56.1 (26-71)
Serum ferritin level (ng/mL)*	
Mean (range)	2175 (1098-4471)
Iron saturation (%) [†]	
Mean (range)	82.5 (60-100)
Liver iron grade	2 (n=5), 3 (n=5), 4 (n=2)

* Normal range: 30-284 ng/mL for males, 6-186 ng/mL for females [†] Normal range: 20%-55%; value available for only nine patients

The quality of sample DNA was unsatisfactory for analysis of the C282Y mutation in one case and the H63D mutation in nine cases. No C282Y mutation was detected in the 49 satisfactory samples. Of the 41 samples that were analysed for H63D mutation, three were heterozygous for H63D and only one was homozygous, giving an H63D allele frequency of 6.1%. One heterozygote had β -thalassaemia major, one had β -thalassaemia minor, and the third had hereditary spherocytosis. The characteristics of these three patients are shown in Table 3. Of the four patients with β -thalassaemia minor, the H63D heterozygote had the highest grade of iron deposition. The only H63D-homozygous patient was a 75-year-old woman with carcinoma of cervix, who died from

Patient	Associated diseases	Sex/age (years)	Haemoglobin level (g/L)	Iron saturation (%)	Serum ferritin level (ng/mL)	Liver iron grade
1	β-Thalassaemia major, hepatitis C virus infection	F/13	99	-	2128	3
2	β-Thalassaemia minor	F/48	95	93	2285	4
3	Hereditary spherocytosis	F/19	125	-	369	2

Table 3. Characteristics of patients who were heterozygous for H63D mutation

pneumonia. A para-mortem liver biopsy showed grade 2 siderosis and mild steatosis; iron overload was not suspected while the patient was alive. None of the 12 idiopathic haemochromatotic patients were positive for either mutation.

Discussion

The discovery of the C282Y mutation in the *HFE* gene has had an enormous impact on the diagnosis of haemochromatosis, because a simple PCR-based genotyping assay can be used to identify most patients definitively.^{18,19} This situation should be contrasted with other genetic hepatic diseases, such as Wilson's disease,²⁰ in which many mutations exist throughout the gene of interest and direct genetic testing is limited to specialist research centres. Furthermore, the method of genetic testing for the mutation is relatively straightforward, because the G \rightarrow A base mutation creates a restriction site for commercially available restriction enzymes. Thus, many hospital laboratories are capable of testing for the mutation.

To the best of our knowledge, this is the first study to analyse HFE mutations in Chinese haemochromatotic patients. The main finding is that the C282Y mutation was not detected in our series of patients. The absence of C282Y mutations implies that the classical form of HLA-linked HHC does not exist in the local population. The association of the C282Y mutation with HHC has been established,²⁻⁸ and the mutation is thought to have arisen in a single individual. approximately 60 to 70 generations ago.²¹ Thus, the global prevalence of this point mutation varies considerably among the populations studied.¹² It is most prevalent in western European populations of Celtic ancestry,²² but fewer patients in Italy and southern France are homozygous for the mutation.^{23,24} In contrast, the C282Y mutation is absent from Asian, African, and Australian indigenous populations.^{12,13,25} The present study confirms the absence of the C282Y mutation among haemochromatotic Chinese patients.

The H63D mutation has been found in diverse ethnic backgrounds and may represent an old mutation.^{12,13} Its clinical significance is less clear than that of the C282Y mutation, and there are conflicting data as to whether it contributes to iron overload in C282Y/H63D compound heterozygotes. The single H63D-homozygous patient in our series had iron overload, but to a degree that was not severe enough to cause symptoms or induce fibrosis.

In this series, the majority of cases of iron overload resulted from secondary causes. Haemochromatosis associated with blood transfusion is easy to diagnose, either from the clinical history or histological material. In contrast, haemochromatosis that is acquired through oral intake-for example, haemoglobinopathies not requiring regular blood transfusion-are difficult to diagnose. The histological appearance of the liver in these patients with siderosis is similar, if not identical, to that in patients with HHC. In Hong Kong, iron overload diseases are uncommon and equipment to measure hepatic iron concentration is not generally available, even in research centres. Using the hepatic iron index, which allows physicians in the West to distinguish between HHC and iron overload due to alcoholism,²⁶ is of little value in this situation. In this study, the cases were associated with haemoglobinopathies, so the degree of iron overload was severe and the indices would have exceeded 1.9. It has been claimed that the minor haemoglobinopathies not requiring regular blood transfusion will not lead to marked iron overload.²⁷ In fact, this is not the case. Nevertheless, severe siderosis has been recently reported in patients with β -thalassaemia minor or haemoglobin H disease and results in liver dysfunction and systemic symptoms such as diabetes.²⁸ Given the large number of thalassaemia carriers in Hong Kong, the occurrence of severe iron overload is probably low. One of the patients in this study had been prescribed iron tablets for more than 10 years, and this may be one of the reasons for such an occurrence. Another aggravating factor, as found in this study, is the presence of the H63D mutation. The allele frequency of this mutation in this series was 6.1%, whereas that of the general population ranges from 2.8% to 3.09%.^{12,13} The mutation seemed to be associated with an erythropoietic abnormality in three of the 16 patients in disease groups 1 and 2.

The main objective of this study was to examine the genetic alterations in a group of Chinese patients with idiopathic haemochromatosis. However, their disease may represent a phenotypically similar but genotypically different heritable trait. Sequencing the coding region of the entire HFE gene in this group of patients may disclose mutations different from the classical ones; such a result has been reported for one non-Caucasian series.²⁹ On the other hand, there are families from southern Italy who have typical HHC without any detectable mutations in the HFE gene.³⁰ The condition has also been demonstrated to be non-HLA-linked in a Chinese woman.³¹ This finding suggests that factors other than mutant HFE genes cause HHC. A study of haemochromatosis in an urban African population has suggested the presence of other types of genetic predisposition to iron overload.³² In addition, Shaheen et al³³ have shown that more than 50% (10/18) of a group of patients with haemochromatosis who lacked the C282Y mutation had other underlying causes of iron overload. In fact, secondary iron overload is the most common cause of haemochromatosis in this locality. The main drawback in this study was the lack of detailed clinical data, the haematological status, or family history of all the patients who were presumed to have primary haemochromatosis. Iron overload as severe as that seen in HHC has been found in patients with alcoholic liver disease, hepatitis C infection, exogenous iron use, or haemoglobinopathies.³⁴ End-stage cirrhosis alone may also cause substantial iron accumulation, regardless of the aetiology of the liver disease.^{35,36} The possibility that the idiopathic haemochromatotic patients in our series had previously unrecognised causes of secondary iron overload cannot be excluded. Prospective studies that carefully exclude secondary causes are thus necessary to identify the cause of disease.

In conclusion, the C282Y mutation is not found in Chinese patients with hepatic iron overload disease. The H63D mutation is found in a minority (9.8%) of patients, in whom it appears to act synergistically with an erythropoietic factor to cause a state of heavy iron overload. For the patients with idiopathic haemo-chromatosis, the possibility of a secondary cause is high, but a genetic basis cannot be excluded. Further studies, such as the sequencing of the coding region of the entire *HFE* gene, are required to further investigate the genetic basis of hepatic iron overload.

Acknowledgement

This study was supported in part by a Hong Kong Polytechnic University Research Grant, No. A/C H-ZJ39.

References

- Edwards CQ, Griffen LM, Goldgar D, Drummond C, Skolnick MH, Kushner JP. Prevalence of haemochromatosis among 11,065 presumably healthy blood donors. N Engl J Med 1988; 318:1355-62.
- Feder JN, Gnirke A, Thomas W, et al. A novel MHC class-I like gene is mutated in patients with hereditary haemochromatosis. Nat Genet 1996;13:399-408.
- 3. Feder JN, Penny DM, Irrinki A, et al. The haemochromatosis gene product complexes with the transferrin receptor and lowers its affinity for ligand binding. Proc Natl Acad Sci USA 1998;95:1472-7.
- Gross CN, Irrinki A, Feder JN, Enns CA. Co-trafficking of HFE, a nonclassical major histocompatibility complex class I protein, with the transferrin receptor implies a role in intracellular iron regulation. J Biol Chem 1998:273:22068-74.
- 5. Jazwinska EC, Cullen LM, Busfield F, et al. Haemochromatosis and *HLA-H*. Nat Genet 1996;14:249-51.
- 6. Jouanolle AM, Gandon G, Jezequel P, et al. Haemochromatosis and *HLA-H*. Nat Genet 1996;14:251-2.
- Beutler E, Gelbart T, West C, et al. Mutation analysis in hereditary haemochromatosis. Blood Cells Mol Dis 1996; 22:187-94.
- Calandro L, Thorsen T, Barcellos L, Griggs J, Baer D, Sensabaugh GE. Mutation analysis in hereditary haemochromatosis [commentary]. Blood Cells Mol Dis 1996;22; 194A-194B.
- 9. Martinez PA, Biron C, Blanc F, et al. Compound heterozygotes for haemochromatosis gene mutations: May they help to understand the pathophysiology of the disease? Blood Cells Mol Dis 1997;23:269-76.
- Sham RL, Ou CY, Cappuccio J, Braggins C, Dunnigan K, Phatak P. Correlation between genotype and phenotype in hereditary haemochromatosis: analysis of 61 cases. Blood Cells Mol Dis 1997;23:314-20.
- Crawford DH, Jazwinska EC, Cullen LM, Powell LW. Expression of *HLA*-linked haemochromatosis in subjects homozygous or heterozygous for the C282Y mutation. Gastroenterology 1998;114:1003-8.
- Merryweather-Clarke AT, Pointon JJ, Shearman JD, Robson KJ. Global prevalence of putative haemochromatosis mutations. J Med Genet 1997;34:275-8.
- Cullen LM, Gao X, Easteal S, Jazwinska EC. The hemochromatosis 845 GÆA and 187 CÆG mutations: prevalence in non-Caucasian populations. Am J Hum Genet 1998;62: 1403-7.
- Scheuer PJ, Williams R, Muir AR. Hepatic pathology in relatives of patients with haemochromatosis. J Pathol Bacteriol 1962;84:53-64.
- Scheuer PJ. Disturbances of copper and iron metabolism. In: Scheuer PJ, Lefkowitch JH, editors. Liver biopsy interpretation. 5th ed. London: WB Saunders; 1994;218-28.
- Mura C, Nousbaum JB, Verger P, et al. Phenotype-genotype correlation in haemochromatosis subjects. Hum Genet 1997; 101:271-6.
- Jouanolle AM, Fergelot P, Gandon G, et al. A candidate gene for hemochromatosis: frequency of the C282Y and H63D mutations. Hum Genet 1997;100:544-7.
- Rossi E, Henderson S, Chin CY, et al. Genotyping as a diagnostic aid in genetic haemochromatosis. J Gastroenterol Hepatol 1999;14:425-8.
- 19. Bacon BR, Olynyk JK, Brunt EM, Britton RS, Wolff RK. *HFE* genotyping in patients with hemochromatosis and other liver

diseases. Ann Intern Med 1999;130:953-62.

- Roberts EA, Cox DW. Wilson disease. In: Arthur MJP, Cleghorn GJ, Creutzfeldt W, Gollan JL, Tytgat GN, editors. Ballière's clinical gastroenterology: international practice and research. London: Ballière Tindall; 1998;237-56.
- Ajioka RS, Jorde LB, Gruen JR, et al. Haplotype analysis of haemochromatosis: Evaluation of different linkage-disequilibrium approaches and evolution of disease chromosomes. Am J Hum Genet 1997;60:1439-47.
- 22. Lucotte G. Celtic origin of the C282Y mutation of haemochromatosis. Blood Cells Mol Dis 1998;24:433-8.
- Carella M, D'Ambrosio L, Totaro A, et al. Mutation analysis of the *HLA-H* gene in Italian haemochromatosis patients. Am J Hum Genet 1997;60:828-32.
- Borot N, Roth M, Malfroy L, et al. Mutations in the MHC class I-like candidate gene for haemochromatosis in French patients. Immunogenetics 1997;45:320-4.
- Roth MP, Giraldo P, Hariti G, et al. Absence of the haemochromatosis gene Cys282Tyr mutation in three ethnic groups from Algeria (MZAB), Ethiopia and Senegal. Immunogenetics 1997;46:222-5.
- Bassett ML, Halliday JW, Powell LW. Value of hepatic tissue iron measurement in early hemochromatosis and determination of the critical iron level associated with fibrosis. Hepatology 1986;6:24-9.
- Bannerman RM, Callender ST, Hardisty RM, Sephton-Smith R. Iron absorption in thalassemia. Br J Haematol 1964;10: 490-5.

- 28. Lin CK, Peng HW, Ho CH, Yung CH. Iron overload in untransfused patients with hemoglobin H disease. Acta Hematol 1990;83:137-9.
- 29. Barton JC, Sawada-Hirai R, Rothenberg BE, Acton RT. Two novel missense mutations of the *HFE* gene (I105T and G93R) and identification of the S65C mutation in Alabama haemochromatosis probands. Blood Cells Mol Dis 1999;25:146-154.
- 30. Pietrangelo A, Montosi G, Totaro A, et al. Hereditary haemochromatosis in adults without pathogenic mutations in the haemochromatosis gene. N Engl J Med 1999;341:725-32.
- Oliver M, Scully L, Guiraudon C, Adams PC. Non-HLA-linked hemochromatosis in a Chinese woman. Dig Dis Sci 1995;40: 1589-91.
- 32. Gangaidzo IT, Moyo VM, Saungweme T, et al. Iron overload in urban Africans in the 1990s. Gut 1999;45:278-83.
- Shaheen NJ, Bacon BR, Grimm IS. Clinical characteristics of hereditary haemochromatosis patients who lack the C282Y mutation. Hepatology 1998;28:526-9.
- Bonkovsky HL, Banner BF, Lambrecht RW, Rubin RB. Iron in liver diseases other than haemochromatosis. Semin Liver Dis 1996;60:65-82.
- Ludwig J, Hashimoto E, Porayko MK, et al. Hemosiderosis in cirrhosis: a study of 447 native livers. Gastroenterology 1997;112:882-8.
- 36. Deugnier Y, Turlin B, Quilleuc D, et al. A reappraisal of hepatic siderosis in patients with end-stage cirrhosis: practical implications for the diagnosis of haemochromatosis. Am J Surg Pathol 1997;21:669-75.